	<b>Standard Operating Procedure</b> <b>Determination of Vitamin K1 by</b> <b>HPLC-UV</b>		<b>SOP Number</b> <b>D-1019</b>	<b>Revision</b> <b>0</b>
			<b>Effective Date</b> <i>05/24/23</i>	<b>Page</b> <b>Page 1 of 6</b>
<b>Written by/ Date</b> <i>CSP 05-22-23</i>		<b>Reviewed by/ Date</b> <i>SAS 05/23/23</i>		<b>Approved by/ Date</b> <i>SSS 05/23/23</i>
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## 1.0 Purpose

This document describes the analytical procedure for the determination of Vitamin K1 in raw materials.

## 2.0 Scope

This procedure applies to the identification and quantification of Vitamin K1 in raw materials. This method was validated under protocol PRTCL-23-0019.

## 3.0 Responsibility

- 3.1 It is the responsibility of QC and Analytical chemists who have verified their ability to execute this procedure to follow this procedure.
- 3.2 It is the responsibility of QC Laboratory Management to implement this procedure and to ensure that the procedure is being followed.
- 3.3 It is the responsibility of QC Laboratory Management and/or Analytical Development Personnel to keep this procedure current with the associated monographs and laboratory practices.

## 4.0 Definitions

- 4.1 **QC** – Quality Control
- 4.2 **ACN** – Acetonitrile
- 4.3 **EtOH** – Ethanol

- 4.4 ACS – American Chemical Society
- 4.5 HPLC – High Performance Liquid Chromatography
- 4.6 UV-Vis – Ultraviolet-Visible (Detection)

## 5.0 References

- 5.1 PRTCL-23-0019, Protocol, Validation of an Analytical Method for the Determination of Vitamin K1 by HPLC-UV

## 6.0 Supplies

- 6.1 Chemicals – All reagents are ACS grade or better
  - 6.1.1 ACN
  - 6.1.2 EtOH
  - 6.1.3 Vitamin K1 (Phytonadione) Reference Standard
- 6.2 Supplies and Glassware **(Use Red Glassware!)**
  - 6.2.1 HPLC vials, 12mm X 32mm with screw cap enclosures w/ septa
  - 6.2.2 **Red** Volumetric glassware
  - 6.2.3 Weigh paper
  - 6.2.4 Syringes with 0.45 $\mu$  nylon syringe filters
- 6.3 Equipment
  - 6.3.1 Suitable gradient HPLC system consisting of a pump, autosampler, column compartment and UV-Vis detector with a chromatographic data handling system
  - 6.3.2 Analytical Balance

6.3.3 Wrist Action Shaker

6.3.4 Sonicator

## 7.0 Procedure

7.1 Mobile Phase and Extraction Solvent / Diluent Preparation

7.1.1 Mobile Phase

7.1.1.1 ACN (Isocratic)

7.1.2 Extraction Solvent / Diluent

7.1.2.1 Extraction Solvent = Diluent = EtOH

7.2 Standard Preparation **(Use Red Glassware!)**

7.2.1 Prepare standard stock at ~0.6 mg/mL analyte in Extraction Solvent. Mechanically shake stock for 15 minutes at ~ ½ volume then QS to volume. Sonicate for 10 minutes then let cool to room temperature. Shake vigorously then dilute stock 2:50 with Diluent. Shake vigorously then filter a 5mL aliquot for analysis, discarding the first 3-4mL of filtrate.

7.3 Sample Preparation **(Use Red Glassware!)**

7.3.1 The validated linear range for the analytical method is 0.0097 – 0.0341 mg/mL.

7.3.2 Extract sufficient sample (based on the raw material manufacturer assay value) with Extraction Solvent in order to generate a sample stock that is ~0.6 mg/mL analyte. Mechanically shake stock for 15 minutes at ~ ½ volume then QS to volume. Sonicate for 10 minutes then let cool to room temperature. Shake vigorously then dilute stock 2:50 with Diluent. Shake vigorously then filter a 5mL aliquot for analysis, discarding the first 3-4mL of filtrate.

7.4 HPLC Parameters

7.4.1 Column: Supelco Ascentis Express RP-Amide, 2.7 $\mu$ m (SPP), 4.6 x 150mm

7.4.2 Column Temperature: 30°C

7.4.3 Flow rate: 1.5 mL/min

7.4.4 Run Time: 10 minutes

7.4.5 Mobile Phase: Isocratic

7.4.6 Wavelength: 248 nm

7.4.7 Injection Volume: 10  $\mu$ L

7.4.8 Suggested 3-D Spectral Range (for Identification): 210nm - 400nm

#### 7.5 Recommended Sequence

7.5.1 Make at least 2 injections of the Diluent.

7.5.2 Make at least five (5) injections of Working Standard.

7.5.3 Make a single injection of each Sample Preparation.

7.5.4 Make a single injection of the Working Standard after every ten (10) sample injections or at the end of the run.

#### 7.6 System Suitability Requirements

7.6.1 The %RSD of five (5) consecutive standard injections is NMT 2%.

7.6.2 The %RSD of all standard injections is NMT 3%.

#### 7.7 Example calculations for determining % Assay:

$$7.7.1 \quad \% = \frac{R_u}{R_s} \times \frac{Wt_{std} \times P}{V_{std}} \times \frac{V_{spl}}{SA} \times 100$$

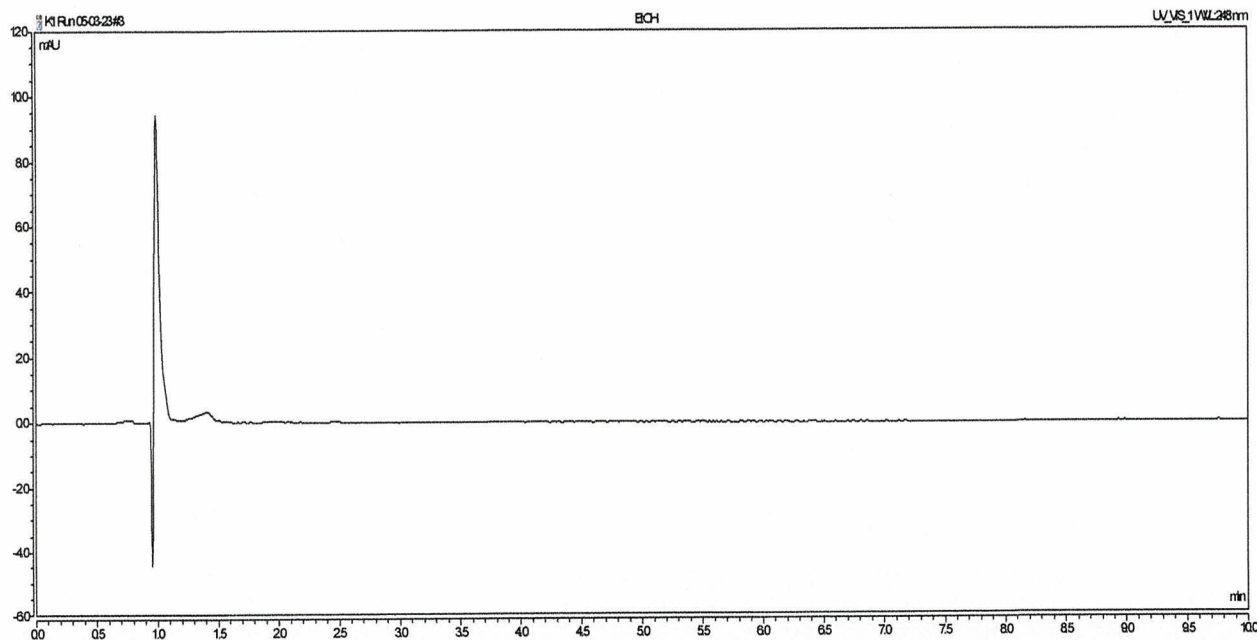
$R_u$	Sample peak area
$R_s$	Mean (All) standard peak area
$Wt_{std}$	Weight of the reference standard, mg
$V_{std}$	Volume of the standard preparation accounting for dilutions, mL
$P$	Purity of the reference standard in decimal format
$SA$	Sample amount, mg
$V_{spl}$	Volume of the sample preparation accounting for dilutions, mL

### 7.8 System Wash, Column Wash and Column Storage

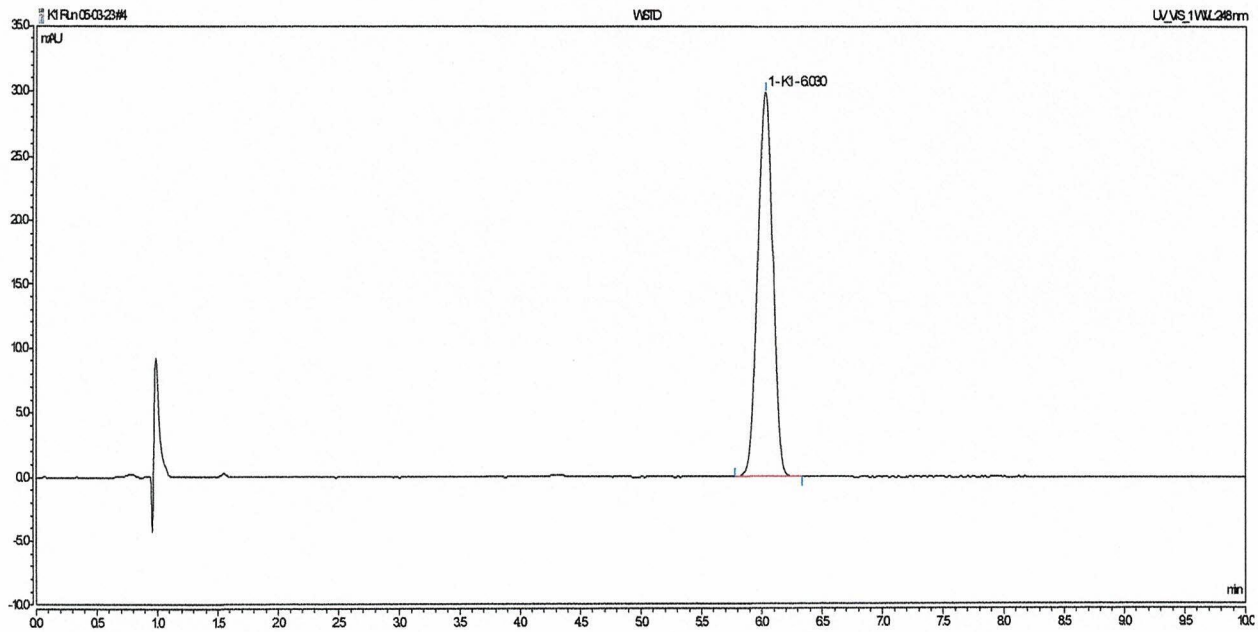
7.8.1 Wash and store the column in 100% ACN.

## 8.0 Chromatograms

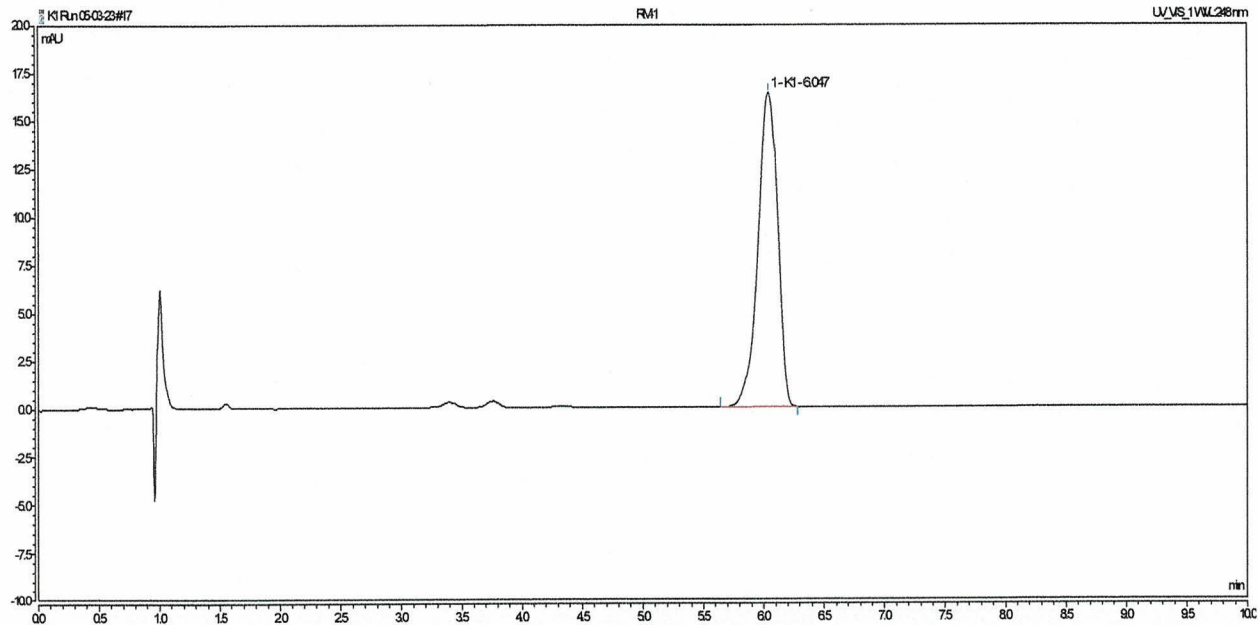
### 8.1 Typical Diluent Chromatogram



8.2 Typical Standard Chromatogram



8.3 Typical Raw Material Chromatogram



9.0 Revision History

Revision	Date	Description of Changes	CCR #	By
0	05/22/23	New procedure.	N/A	C. Perry